ROBL-CRG	Experin Interac produc related absorp	ment title: tion of uraniur ed by soil bac model compo tion spectrosc	Experiment number: 20-01-661	
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Report: Synchrotron-based EXAFS spectroscopy is a powerful technique to obtain structural information on radionuclide bioligand species in solution. As an example pyoverdin-type siderophores are a unique class of bioligands, with a high potential to dissolve, bind, and thus transport uranium in the environment. Pyoverdins are secreted from fluorescent *Pseudomonas* species which are ubiquitous soil bacteria. The aim of this study is to explore structural parameter of soluble U(VI) species with pyoverdins and related model compounds. This includes also relevant model systems for the cell envelope of bacteria. The present report is focussed on the first topic: soluble U(VI) bioligand species and their structural characteristics.



Fig. 1: U L_{III} -edge k^3 -weighted EXAFS spectra (left) and the corresponding Fourier transforms (right) and the theoretical fits (red line).

Experimental. U L_{III}-edge EXAFS measurements were carried out with test solutions containing 5×10^{-4} or 0.001 M UO₂²⁺ and pyoverdins (PYO) or related model compounds at an ionic strength of 0.1 M NaClO₄. The pH was varied between 2 and 8 depending on the bioligand. The samples were measured at room temperature either in fluorescence or in transmission mode. The model compounds simulate the hydroxamate function [salicylhydroxamic acid (SHA), benzohydroxamic acid (BHA) and desferrioxamine B (DFO)] and the chromophore [2,3-dihydroxynaphthalene (NAP)] of the pyoverdin molecule.

Results. A selection of the measured EXAFS oscillations and corresponding Fourier transforms are presented in Fig. 1. The extracted structural parameters are summarized in Table 1.

$ \begin{array}{c ccccccccccccccccccccccccccccccccccc$	Sample	Shell	Ν	R (A)	σ² (A²)	∆E₀ (eV)
$\begin{array}{c c c c c c c c c c c c c c c c c c c $	A	U-O _{axial}	2f	1.76 ₆	0.0016	10.5
$\begin{array}{c ccccccccccccccccccccccccccccccccccc$		U-O _{equatorial}	5.0	2.41 ₈	0.0068	
$\begin{array}{c c c c c c c c c c c c c c c c c c c $	B (SHA)	U-O _{axial}	2f	1.77 ₅	0.0026	13.4
$\begin{array}{cccccccccccccccccccccccccccccccccccc$		U-O _{equatorial}	4.7	2.41 ₅	0.0063	
$\begin{array}{c c c c c c c c c c c c c c c c c c c $	C (BHA)	U-O _{axial}	2f	1.77 ₃	0.0027	12.3
$\begin{array}{c ccccccccccccccccccccccccccccccccccc$		U-O _{equatorial}	4.8	2.40 ₂	0.0064	
$\begin{array}{c ccccccccccccccccccccccccccccccccccc$	D (DFO)	U-O _{axial}	2f	1.78 ₁	0.0023	10.3
$\begin{array}{c ccccccccccccccccccccccccccccccccccc$	pH 3.2	U-O _{equatorial}	4.9	2.40 ₀	0.0070	
$\begin{array}{c ccccccccccccccccccccccccccccccccccc$		U-C/N	2f	3.22	0.0040	
$\begin{array}{c ccccccccccccccccccccccccccccccccccc$	E (DFO)	U-O _{axial}	2f	1.78₅	0.0024	9.7
$\begin{array}{c ccccccccccccccccccccccccccccccccccc$	pH 4.0	U-O _{equatorial}	4.8	2.38 7	0.0069	
$\begin{array}{cccccccccccccccccccccccccccccccccccc$		U-C/N	2f	3.22	0.0015	
$\begin{array}{c ccccccccccccccccccccccccccccccccccc$	F (NAP)	U-O _{axial}	2f	1.76 ₈	0.0017	18.6
$\begin{array}{cccccccccccccccccccccccccccccccccccc$	pH 3.5	U-O _{equatorial}	5.6	2.40 ₅	0.0078	
$\begin{array}{c cccccc} pH 8.3 & U-O_{equatorial} & 6.3 & \textbf{2.36}_9 & 0.0095 \\ \hline H (PYO) & U-O_{axial} & 2f & \textbf{1.78}_8 & 0.0023 & \textbf{12.8} \\ pH 6.0 & U-O_{equatorial} & 6f & \textbf{2.35}_3 & 0.0107 \\ U-C/N & 2f & \textbf{2.89} & 0.0047 \\ \hline \\ U(VI)-PCA^{[1]} & U-O_{axial} & 2f & \textbf{1.81} & 0.0013 & -14 \\ pH 10.0 & U-O_{equatorial} & 5.8 & \textbf{2.37}_4 & 0.0071 \\ \hline \end{array}$	G (NAP)	U-O _{axial}	2f	1.79 ₇	0.0018	18.3
$\begin{array}{c ccccccccccccccccccccccccccccccccccc$	pH 8.3	U-O _{equatorial}	6.3	2.36 ₉	0.0095	
$\begin{array}{ccccccc} H \left(PYO \right) & U-O_{axial} & 2f & 1.78_8 & 0.0023 & 12.8 \\ pH 6.0 & U-O_{equatorial} & 6f & 2.35_3 & 0.0107 \\ U-C/N & 2f & 2.89 & 0.0047 \\ \end{array}$						
$\begin{array}{c ccccccccccccccccccccccccccccccccccc$	H (PYO)	U-O _{axial}	2f	1.78 ₈	0.0023	12.8
U-C/N 2f 2.89 0.0047 U(VI)-PCA ^[1] U-O _{axial} 2f 1.81 0.0013 -14 pH 10.0 U-O _{equatorial} 5.8 2.37 4 0.0071 -14	pH 6.0	U-O _{equatorial}	6f	2.35 ₃	0.0107	
U(VI)-PCA ^[1] U-O _{axial} 2f 1.81 0.0013 -14 pH 10.0 U-O _{equatorial} 5.8 2.37 ₄ 0.0071	-	U-C/N	2f	2.89	0.0047	
U(VI)-PCA ^[1] U-O _{axial} 2f 1.81 0.0013 -14 pH 10.0 U-O _{equatorial} 5.8 2.37 ₄ 0.0071						
pH 10.0 U-O _{equatorial} 5.8 2.37 ₄ 0.0071	U(VI)-PCA ^[1]	U-O _{axial}	2f	1.81	0.0013	-14
	pH 10.0	U-O _{equatorial}	5.8	2.37 ₄	0.0071	

Table 1: Summary of the determined structural parameters.

f: fixed during the fit.

The differences in the EXAFS oscillations of the bioligand containing samples B to H compared to the free uranyl ion (sample A) within a k-range between 6 and 9 Å⁻¹ clearly indicates the complexation of U(VI). These U(VI)-species are characterized by both a lengthening of the U-O_{ax} and a shortening of the U-O_{eq} distance with increasing pH. The structural parameters of the U(VI)-pyoverdin sample shows strong similarities with those of 1:1 complexes of U(VI) with protocatechuic acid and catechol [1]. Hence there is a strong affinity of U(VI) to the catechol functionality of the pyoverdin molecule. However, the coordination of U(VI) to hydroxamate groups (samples B to E) results also in a shortening of the U-O_{eq} distance.

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/1/ Roßberg, A. et al. (2000) Radiochim. Acta 88, 593-597.