

In most cases two or more measurements were performed for each sample, in particular for fluorescence detection. The S/N ratio for XANES was good on both type of samples (filters and liquid cells). EXAFS measurements were characterized by some noise and irreproducibilities due to the instability of the old BM08-Lisa monochromator. Absolute energy scale of the spectra was set by comparison with the Se edge which was simultaneously measured at each time on a metallic selenium foil placed after the samples.

Main Results:

Preliminary data processing and analysis were performed using the ATHENA Data Processing software (<https://bruceravel.github.io/demeter/>) after alignment of the XAS spectra.

We found that inside the bacterial cells (millipore filters samples) intermediate oxidation states between selenite (4+) and elemental selenium (0) are present in the very early stages of incubation and that already after 0.5 hour elemental selenium neatly prevails (Fig.1, left side); while in the suspension at least up to 12/16 hours of incubation the main component is selenite (Fig.1, right side), thus indicating that the original selenite content has not been fully reduced.

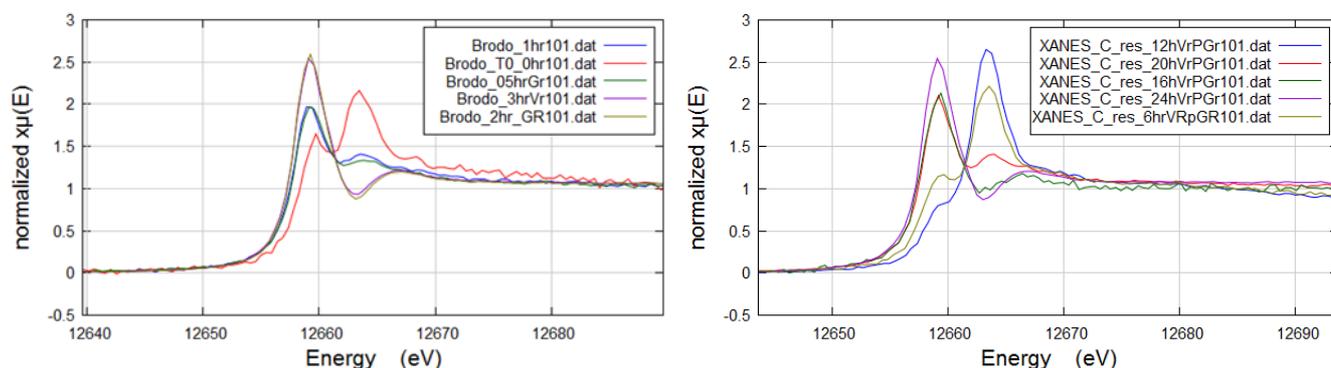


Fig. 1. On the left: XANES on dry bacterial cells of *Stenotrophomonas maltophilia* SeITE02 grown in nutrient broth after addition of 0.5 mM Na_2SeO_3 at different incubation times: from 0 to 3 hrs. On the right: XANES on bacterial cells as suspension, grown in nutrient broth after incubation from 6 to 24 hrs.

The expected role of the cytoplasm in selenite reduction was confirmed. Moreover, we found that the process is slower in the extracted proteic fraction than in the cytoplasm inside the cell. The reduction of selenite by the proteic fraction occurs between 12 and 24 hours (Fig.2, right side), while it appears earlier, between 6 and 12 hours, and is completed after 12 hours, in the cytoplasm purified from the bacterial cells grown with selenite (Fig. 2, left side), suggesting that different mechanisms of selenite reduction could be involved inside the cells.

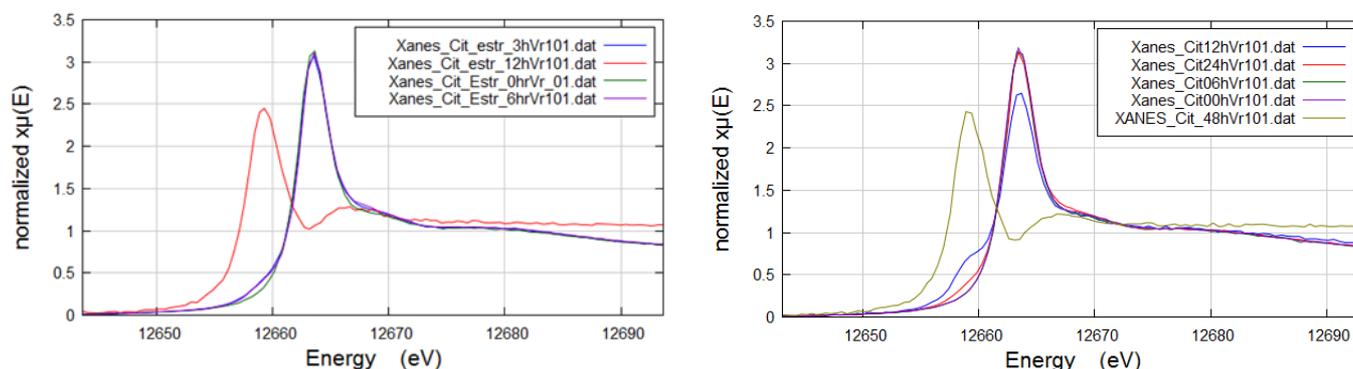


Fig. 2. On the left: XANES on the cytoplasm purified from *Stenotrophomonas maltophilia* SeITE02 grown in nutrient broth with 0.5 mM Na_2SeO_3 after 0, 3, 6, 12 hrs of incubation time. On the right: XANES on the cytoplasm protein fractions, obtained from *Stenotrophomonas maltophilia* SeITE02 grown in nutrient broth for 24 hrs, exposed to 0.5 mM Na_2SeO_3 for 0, 6, 12, 24, 48 hrs.

As regards SeNPs, the absorption edge resulted to be always that of red selenium, independently from the sonication process, indicating that the transition to the grey form seen in the previous experiment is due to the spontaneous transition of Se to the more stable grey form. Moreover, EXAFS measurements at liquid nitrogen temperature on the home-prepared red selenium and on the SeNPs show that there is no long range order in red selenium in any sample, hence suggesting that red selenium is always in the amorphous form.